

**DEVELOPMENT AND APPLICATION OF REAL-TIME PCR  
METHODS FOR THE DETECTION AND TYPING OF  
EXTENDED-SPECTRUM  $\beta$ -LACTAMASES IN CLINICAL  
ISOLATES OF *ENTEROBACTERIACEAE***

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**РАЗРАБОТКА И ПРИМЕНЕНИЕ МЕТОДОВ ПЦР В  
РЕЖИМЕ РЕАЛЬНОГО ВРЕМЕНИ ДЛЯ ИДЕНТИФИКАЦИИ  
 $\beta$ -ЛАКТАМАЗ РАСШИРЕННОГО СПЕКТРА У  
КЛИНИЧЕСКИХ ШТАММОВ ЭНТЕРОБАКТЕРИЙ**

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# EXTENDED-SPECTRUM $\beta$ -LACTAMASES (ESBLs)

- Active against all  $\beta$ -lactams (penicillins, I-IV gen. cephalosporins, aztreonam) except cephamycins and carbapenems
- (- -) Inhibited by active-site directed inhibitors (clavulanic acid, sulbactam, tazobactam)
- Ambler molecular class **A** or class **D** (active site serine  $\beta$ -lactamases)
- Bush, Jacoby and Medeiros functional groups: **2be**, **2d**, **2f**
- Multiple genetic types: **CTX-M**, **SHV**, **TEM**, etc.
- Typically plasmid-encoded, often associated with mobile elements
- Produced mainly by *Enterobacteriaceae*

# WHY ESBLs ARE IMPORTANT?

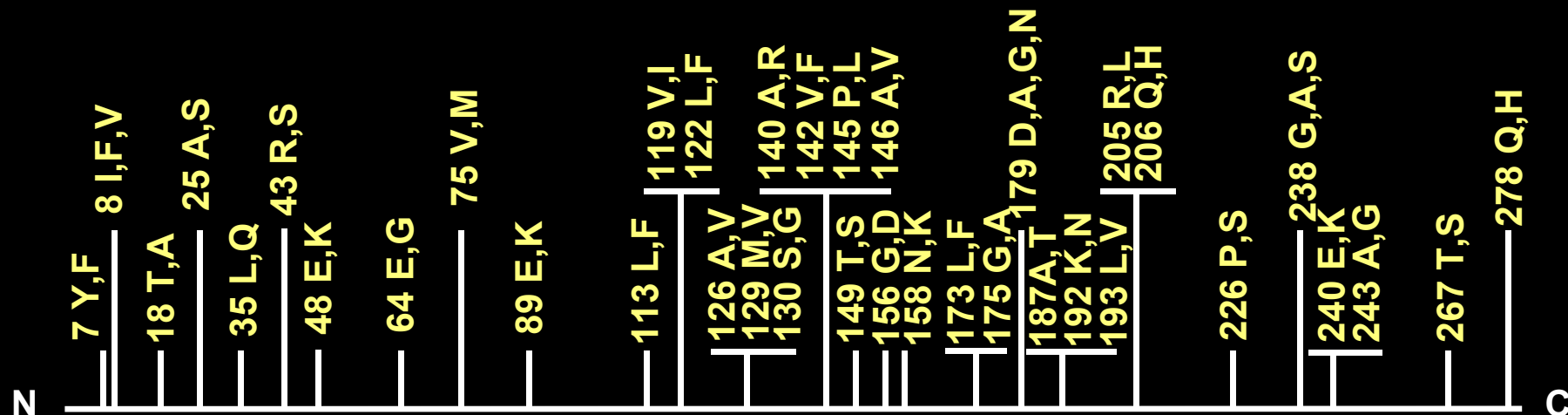
- **Confer resistance to the most commonly used antibiotics** which constitute the first-line therapy for nosocomial infections
- **Difficult to detect by routine susceptibility tests:**  
**FALSE** susceptible ESBL producers are a **common cause of treatment failures with modern cephalosporins**
- **Complex epidemiology:**  
**may rapidly spread by clonal transmission or plasmid transfer**
- **Frequently associated with resistance determinants to non-β-lactam agents (fluoroquinolones, aminoglycosides, tetracyclines, sulfonamides, etc).**  
**ESBL producers are often multiply resistant**

# MOLECULAR TYPING OF ESBLs

- Provides a basis for understanding the epidemiology and evolution of ESBLs
- There is no one single molecular test to detect all ESBL types
- Numerous methods have been proposed for **rapid identification of ESBLs** that belong to certain genetic groups (e.g. TEM, SHV, CTX-M):
  - Spoligotyping;
  - PCR-RFLP;
  - PCR-SSCP;
  - LCR;
  - minisequencing;
  - real-time PCR
- But **none of these techniques can identify all members of any group** in a single step

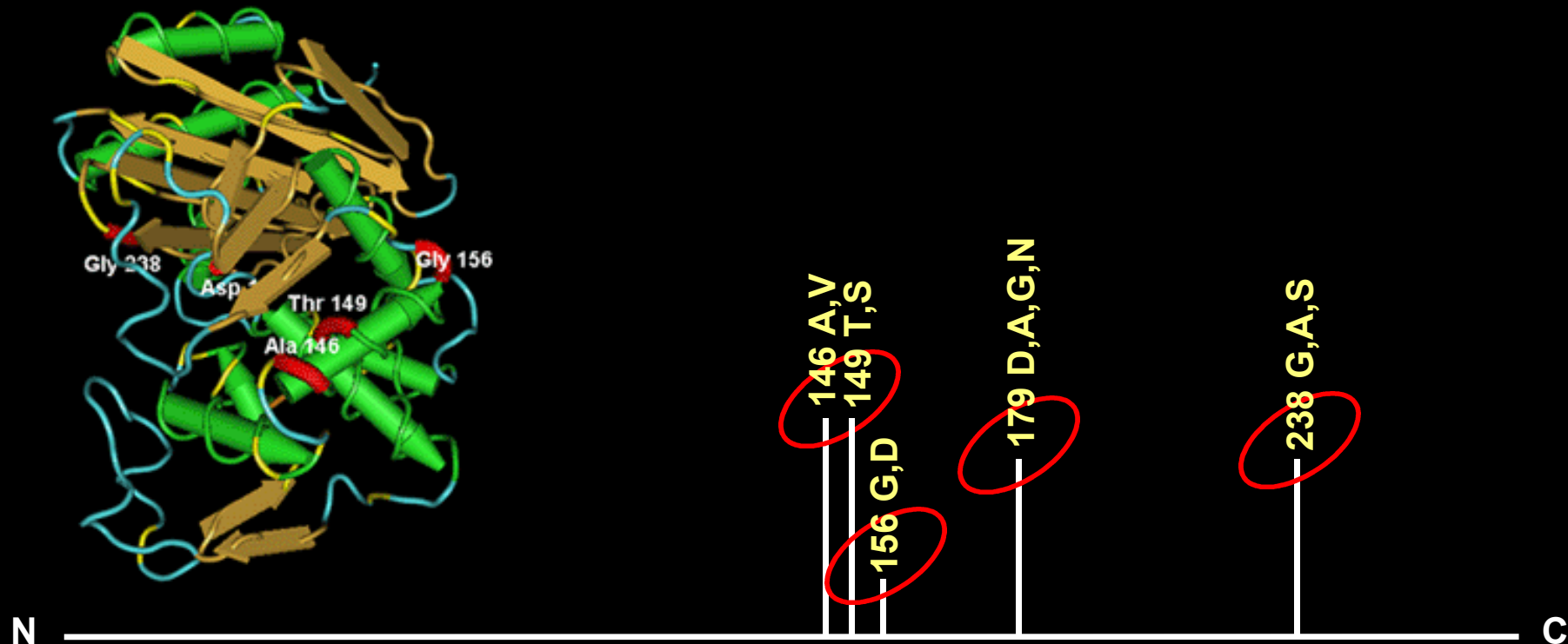
# THE SHV-TYPE ESBLs

- SHV-1 penicillinase (»32 kDa) is a direct progenitor
  - species-specific (chromosomally encoded) in *Klebsiella pneumoniae*
  - often plasmid mediated in different species of *Enterobacteriaceae*
- SHV ESBLs evolve by acquisition of point mutations
- Over 60 amino acid variants currently exist\*



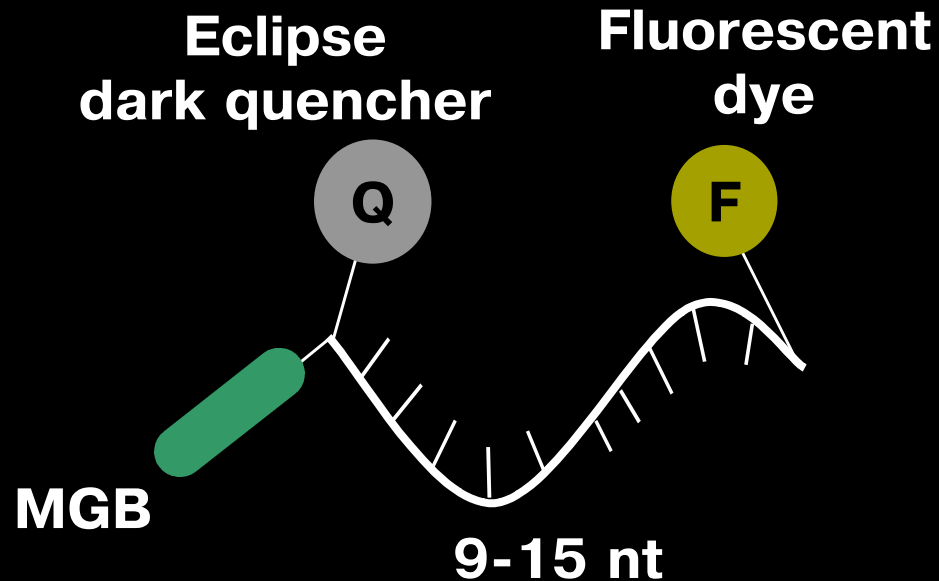
\* <http://www.lahey.org/studies/webt.htm>

# THE SHV-TYPE ESBLs



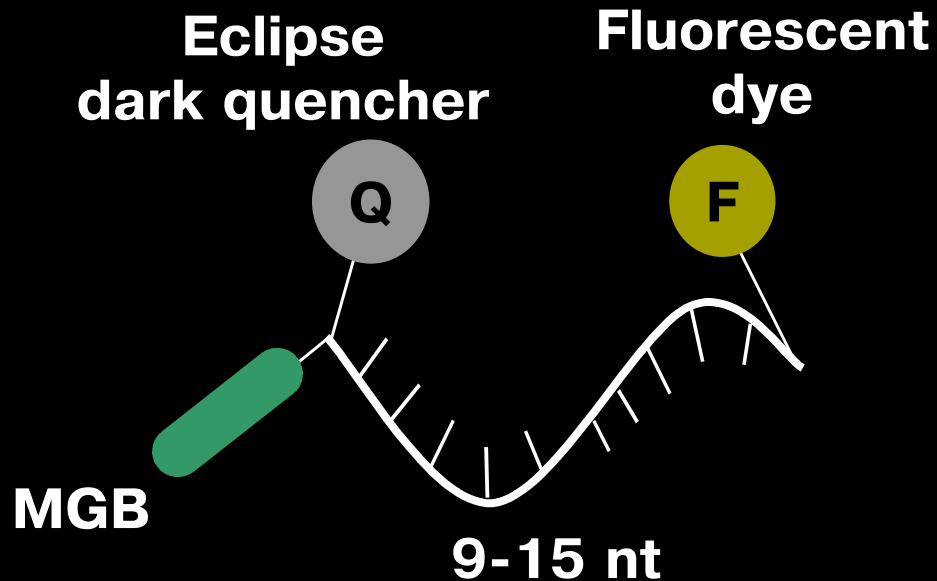
Substitutions at only a few amino acid residues are required for extended-spectrum activity

# MGB ECLIPSE™ PROBES\*



- **Contain minor groove binder (MGB) and Eclipse dark quencher at 5' end**
- **Contain fluorescent dye at 3' end**
- **Short probes (9-15 nt) with high  $T_m$  and precise binding to target**

# MGB ECLIPSE™ PROBES\*

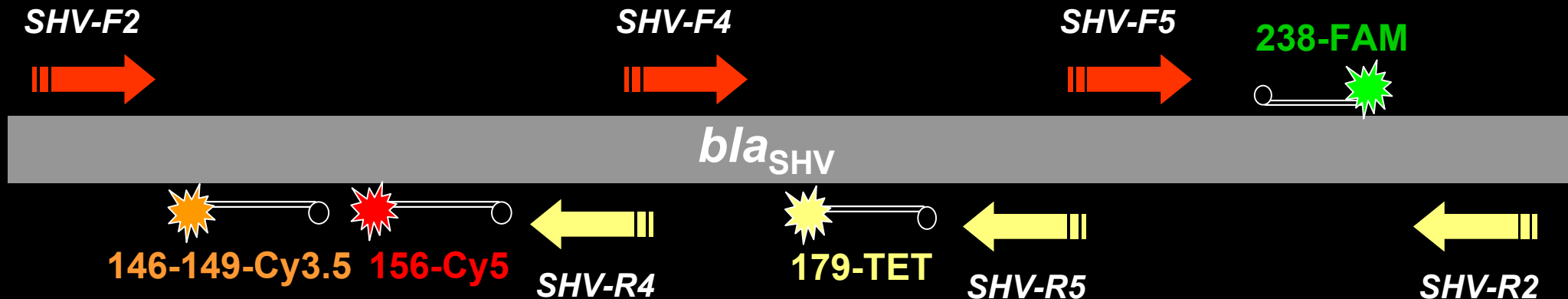


- Quenched in random coil state
- Fluorescing when bond to target
- 5'-nuclease resistant  $\Psi$  suitable for postamplification meting curve analysis





# DESIGN OF MULTIPLEX SINGLE-TUBE PCR AND MELTING CURVE ANALYSIS FOR DETECTION OF SHV ESBLs

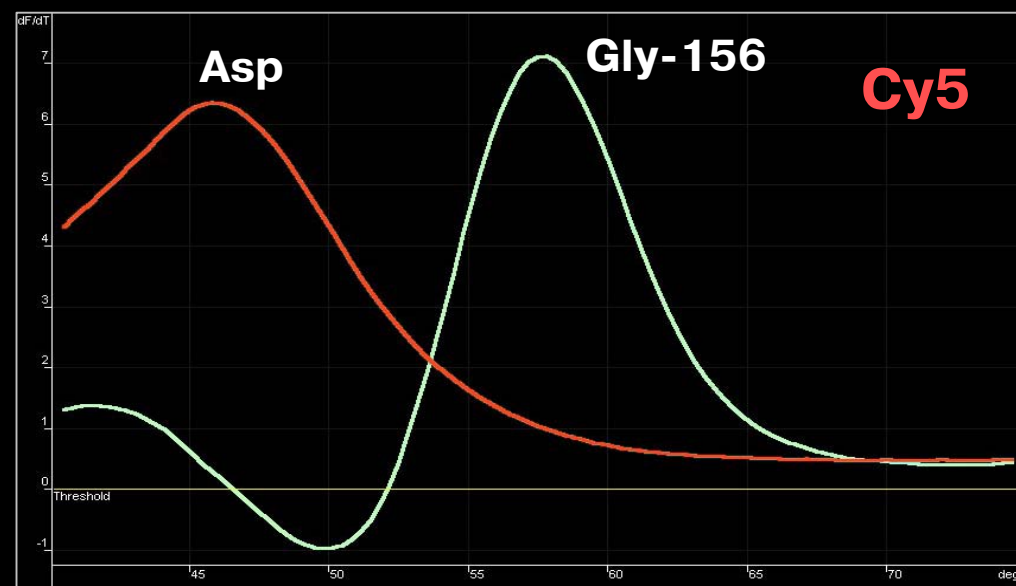
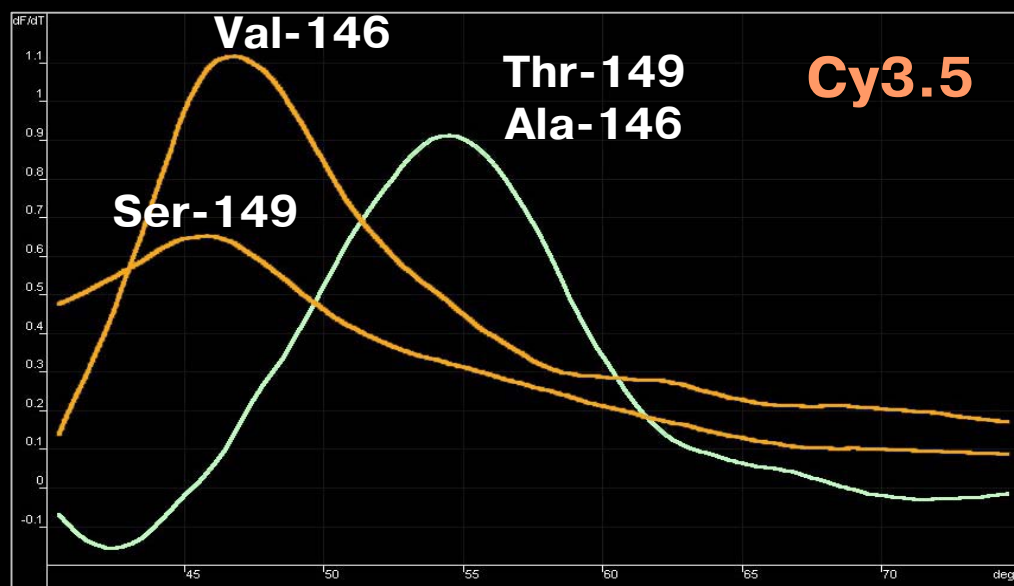
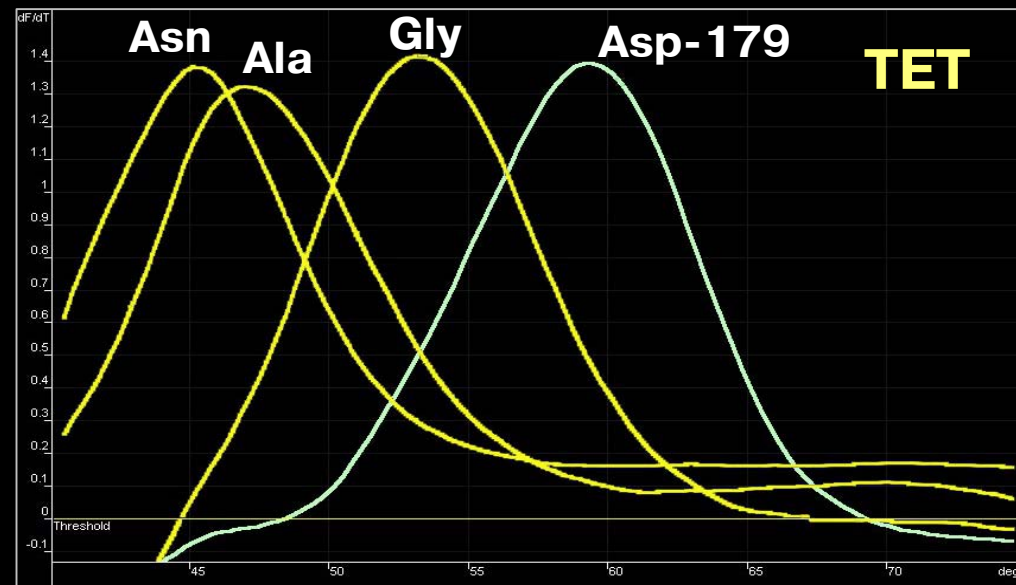


- 4 differentially labelled probes complementary to WT sequences at key mutation sites
- Each mutation specifically shifts the  $T_m$  of the corresponding probe
- Asymmetric primer ratio to favour formation of the strands complementary to the probes

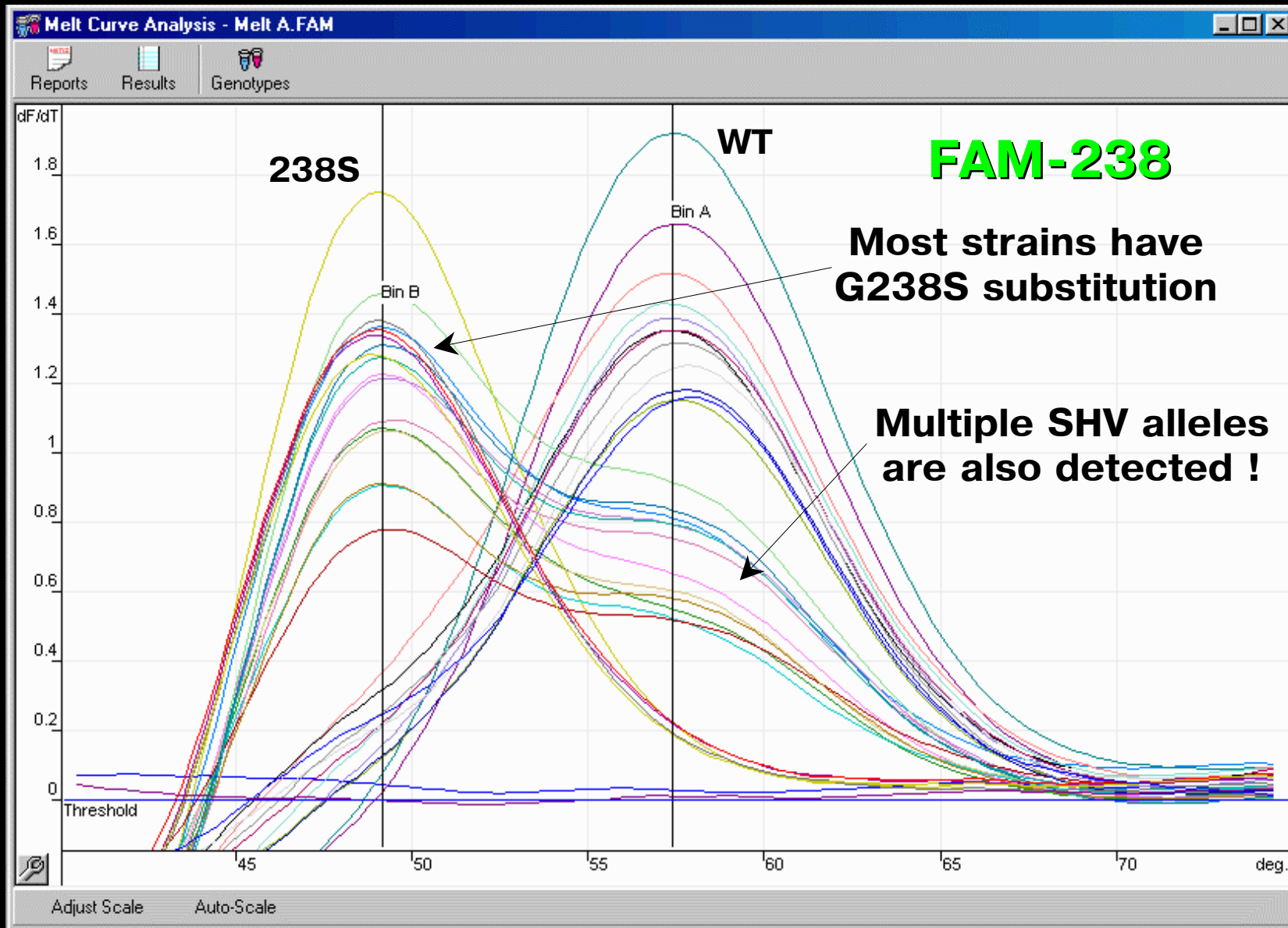
# **CONTROL STRAINS WITH KNOWN MUTATIONS IN SHVs**

<b>Mutation</b>	<b>Detected with probe</b>
<b>WT, non-ESBL control (SHV-1)</b>	<b>All probes</b>
<b>Ser-238 (SHV-2, -3, -4)</b>	<b>238-FAM</b>
<b>Ser-238 + Lys-240 (SHV-5)</b>	<b>238-FAM</b>
<b>Ala-238 (SHV-18)</b>	<b>238-FAM</b>
<b>Ala-179 (SHV-6)</b>	<b>179-TET</b>
<b>Asn-179 (SHV-8)</b>	<b>179-TET</b>
<b>Gly-179 (site-directed mut.)</b>	<b>179-TET</b>
<b>Asp-156 (site-directed mut.)</b>	<b>156-Cy3.5</b>
<b>Ser-149 (site-directed mut.)</b>	<b>146-149-Cy5</b>
<b>Val-146 (site-directed mut.)</b>	<b>146-149-Cy5</b>

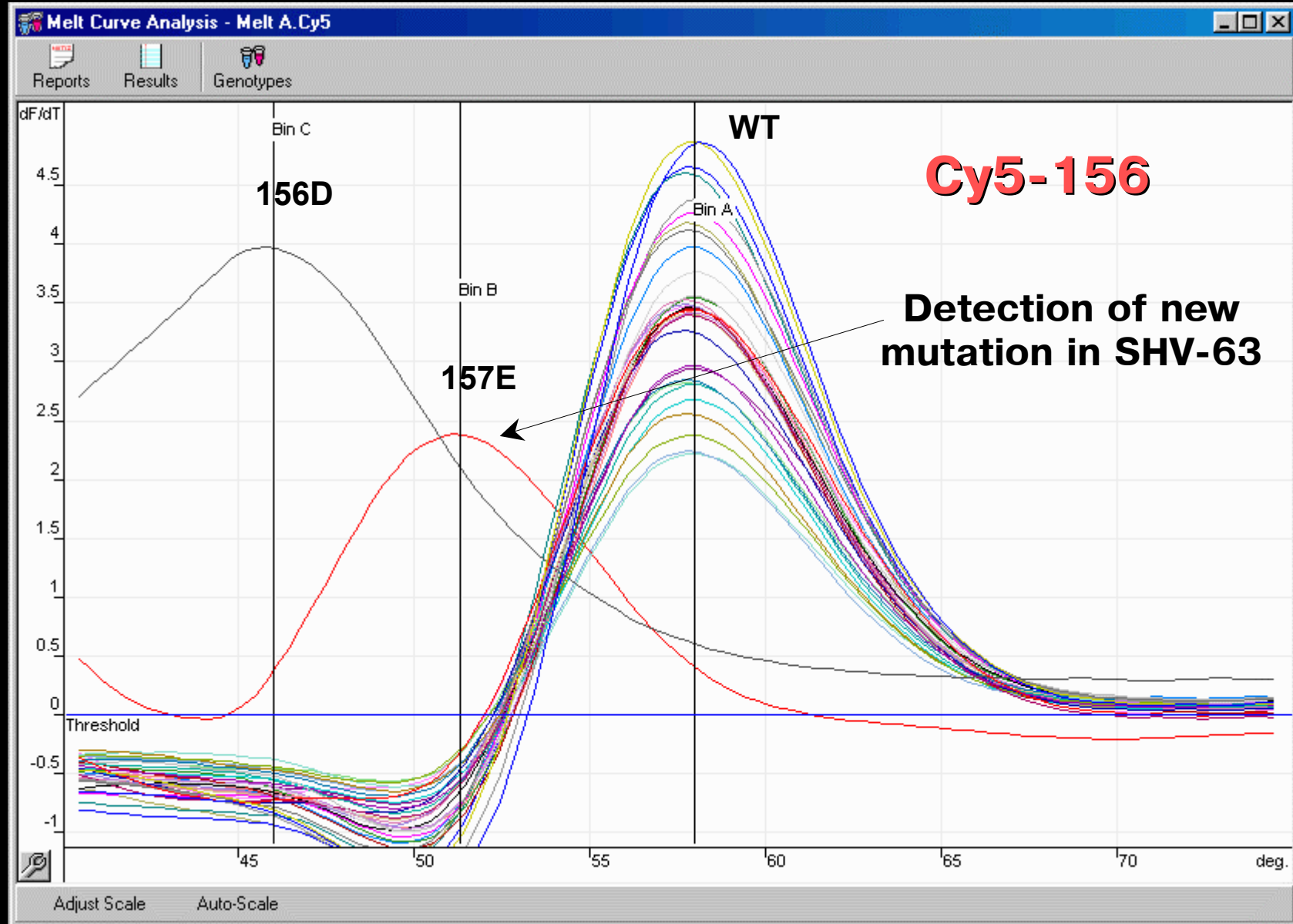
# DETECTION AND DIFFERENTIATION OF ALL THE KNOWN ESBL MUTATIONS BY MELTING CURVE ANALYSIS



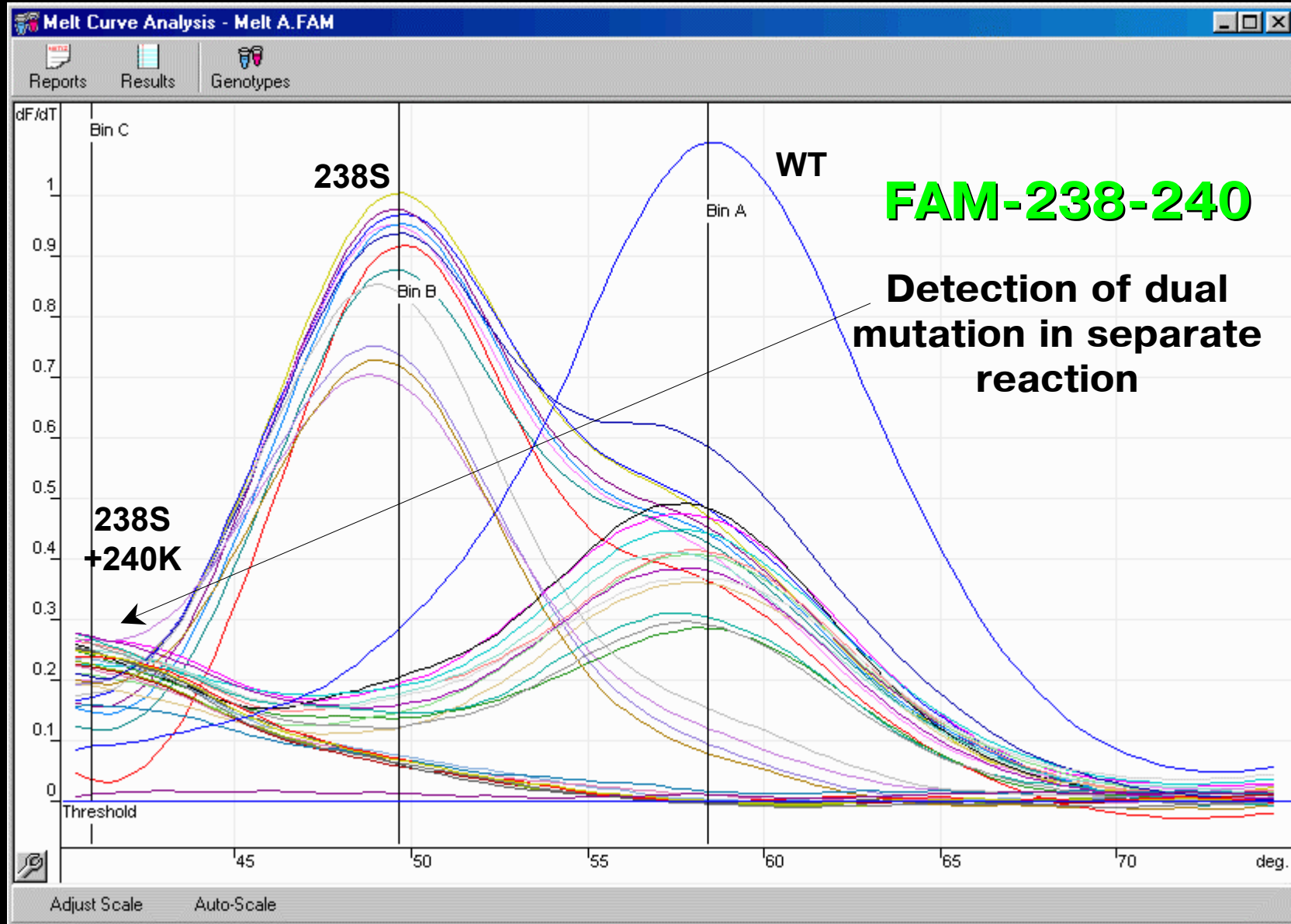
# ANALYSIS OF CLINICAL ISOLATES



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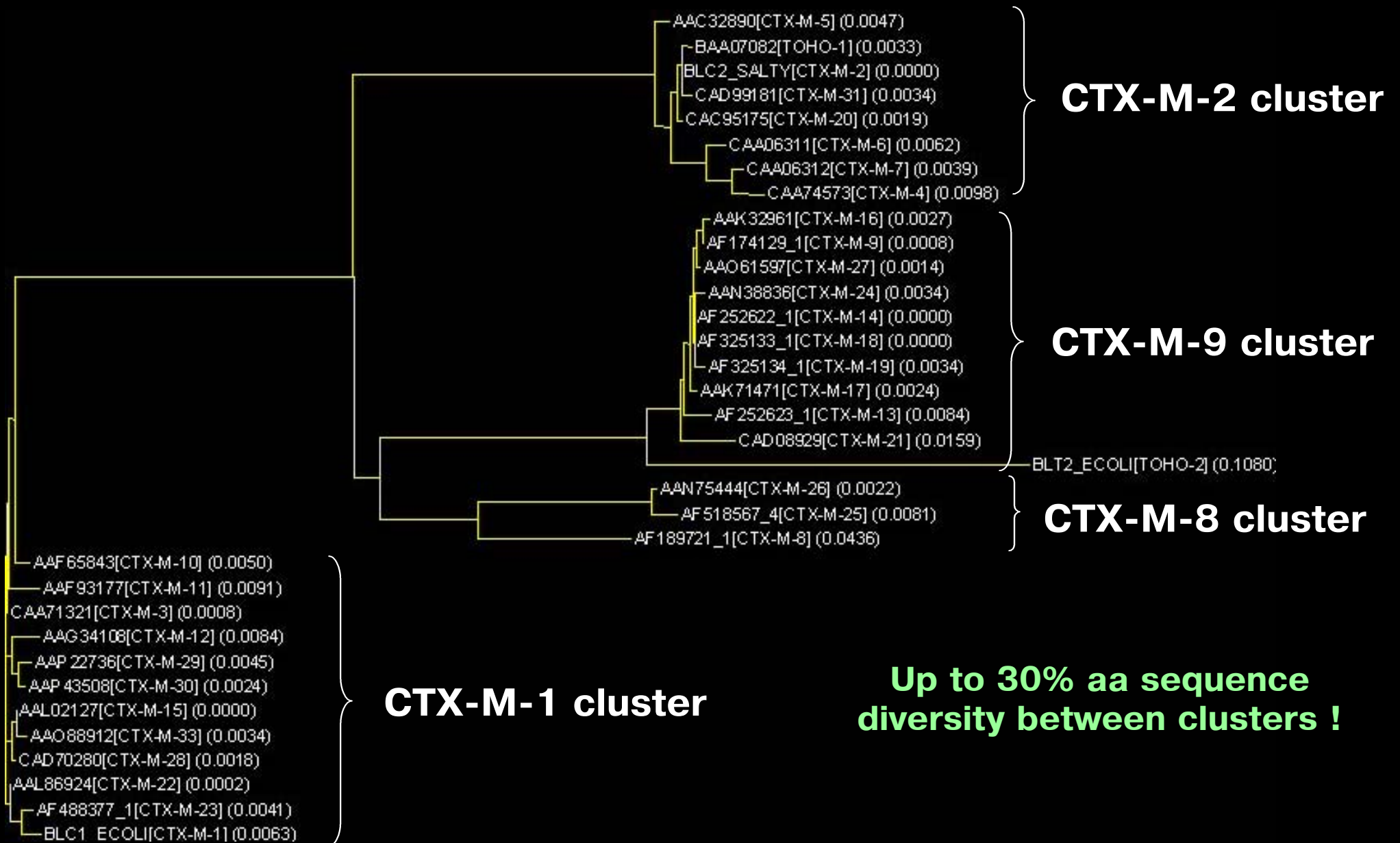


# **DETECTION OF SHV ESBLs BY MULTIPLEX REAL-TIME PCR WITH MGB ECLIPSE™ PROBES**

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- **All the known mutations conferring extended-spectrum activity on SHV  $\beta$ -lactamases can be detected and discriminated in a single reaction**
- **To our knowledge, this is the first example of detection of 8 mutations in 5 codons using a single-tube real-time PCR**
- **New mutations at the key codons can also be identified**
- **Plasmid-mediated SHV ESBLs can be detected on the background of chromosomally-encoded SHV-1 in *K.pneumoniae***
- **Extremely fast and processive  
( $< 4$  h for DNA isolation, PCR and melting curve analysis with 33 strains without opening the tube)**
- **No risk of contamination by PCR products**

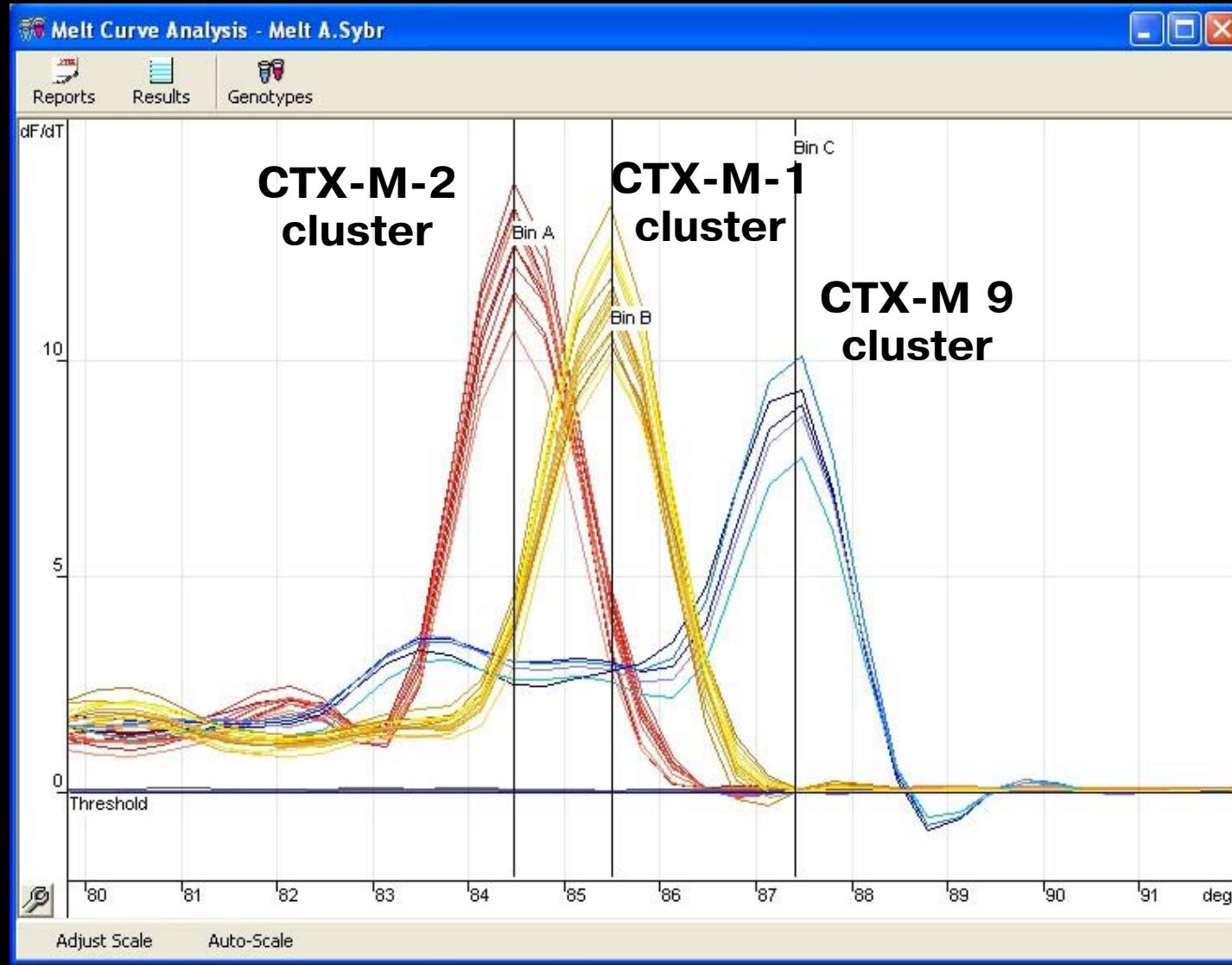
# THE CTX-M-TYPE ESBLs



**Up to 30% aa sequence diversity between clusters !**



# REAL-TIME PCR WITH UNIVERSAL CTX-M PRIMERS AND SYBR GREEN I



- Primers were designed to amplify 519bp PCR-product from all the subtypes
- Members of different clusters can be distinguished by melting curve analysis

# INCIDENCE OF ESBL-PRODUCING *ENTEROBACTERIACEAE* IN RUSSIAN ICUs

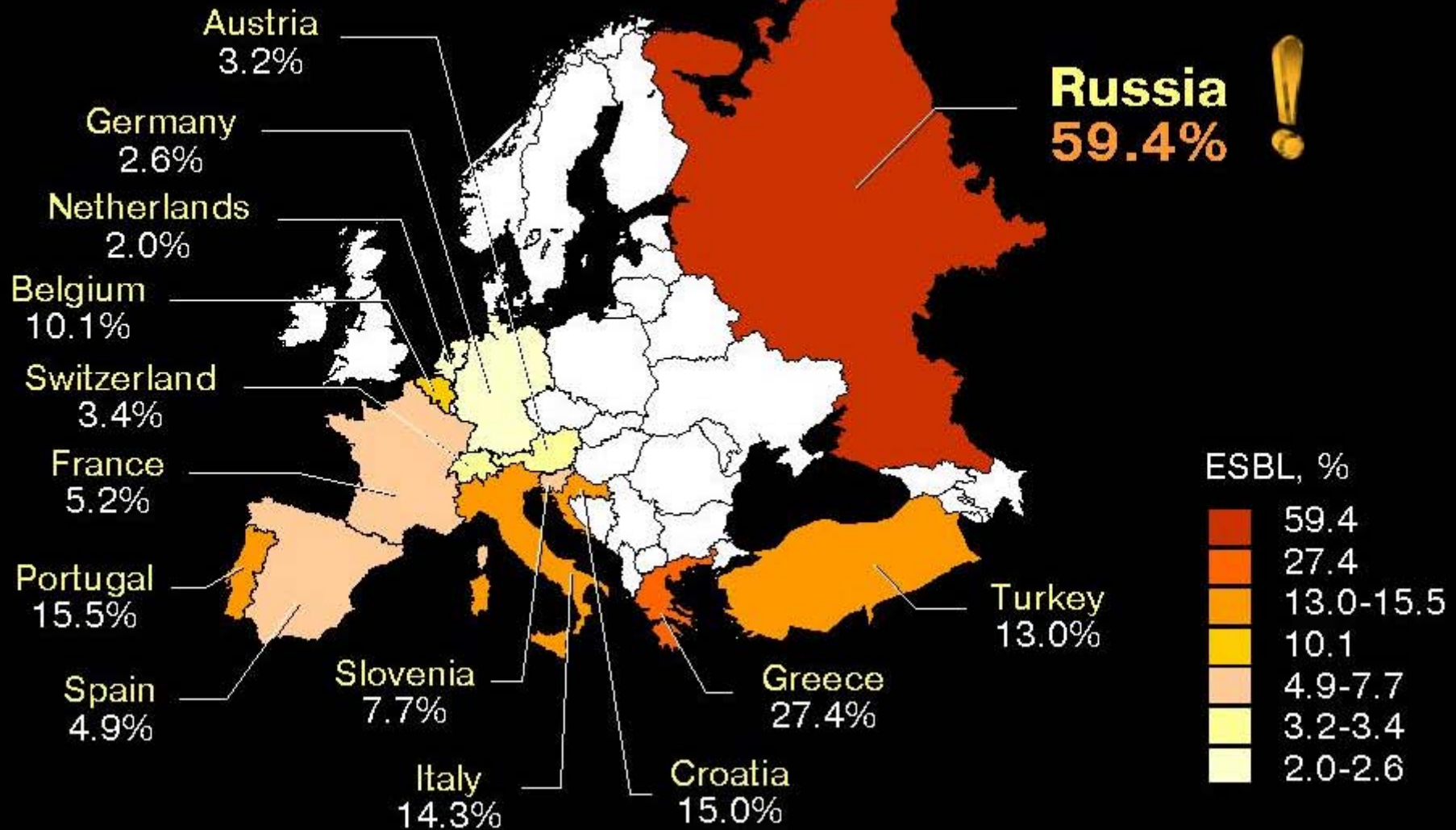
Data from RESORT study - 2003  
648 strains from 33 hospitals of 22 cities



# INCIDENCE OF ESBL PRODUCTION IN NOSOCOMIAL ENTEROBACTERIACEAE

Europe – PEARLS Study 2001–2002

Russia – RESORT Study 2003



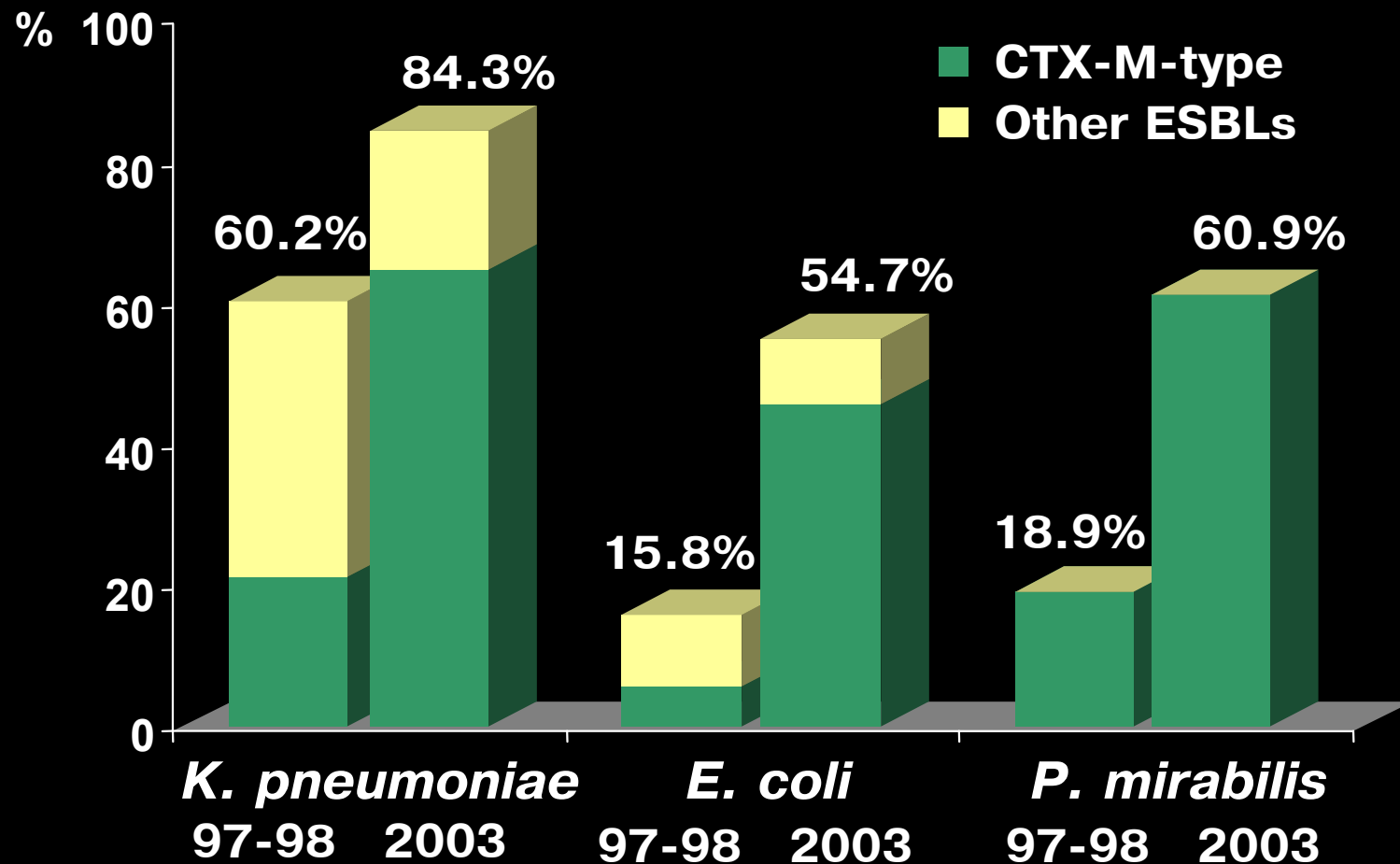
\* S.K. Bouchillon et al., IJAA 2004 (24): 119–24

\*\*M.Edelstein et al., ICAAC, 2004, P.C2-1331

# DRAMATIC INCREASE IN THE PROPORTION OF CTX-M ESBLs IN RUSSIA

1998-99 – NPRS study\*

2003 – RESORT study\*\*

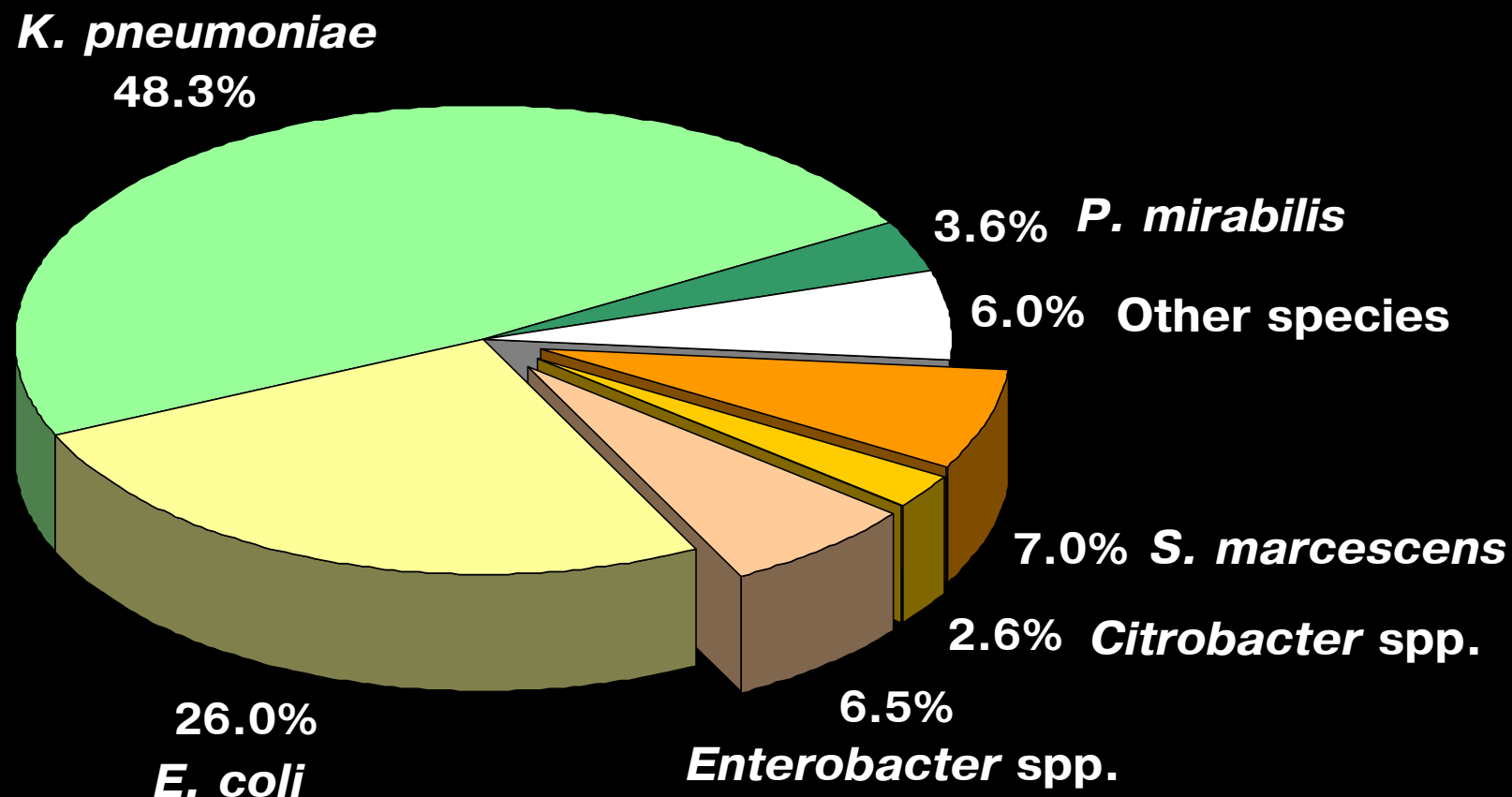


\* M.Edelstein et al., AAC 2003 (47): 3724-32

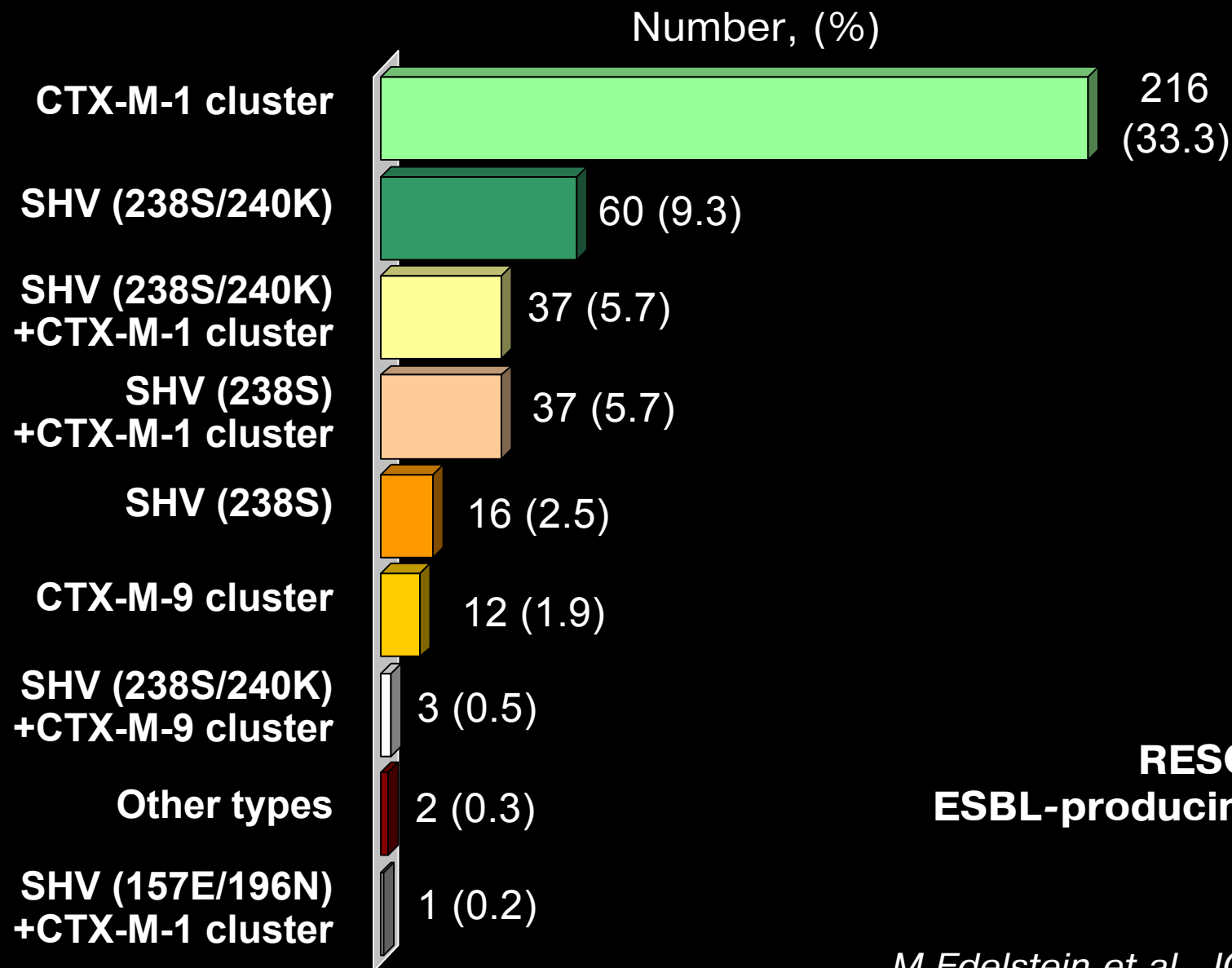
\*\* M.Edelstein et al., ICAAC, 2004, P.C2-1331

# **SPECIES DISTRIBUTION AMONG ESBL-PRODUCING *ENTEROBACTERIACEAE***

**RESORT study – 2003  
ESBL-producing strains (n=384)**



# PREVALENCES OF DIFFERENT ESBL TYPES AND THEIR COMBINATIONS IN RUSSIAN NOSOCOMIAL ISOLATES



**RESORT Study – 2003**  
**ESBL-producing strains (n=385)**

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